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(54) Title: PLANT PROMOTER

(57) Abstract

A plant promoter is provided, which was identified as a promoter for two proline-rich protein (PRP) genes. The promoter can be used to control the expression of genes in plant tissues.

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PLANT PROMOTER

The present invention relates to a plant promoter.

The major structural components of the plant cell wall in terms of quantity are polysaccharides, including cellulose, hemicellulose and pectin polymers which in certain specialised cell types, e.g. vascular/epidermal cells, are complemented by further components whose presence is adapted to the functional role of the cell. The plant cell wall of dicotyledonous plants has also been shown to contain two types of structural protein, the hydroxyproline-rich glycoproteins (HGRPs) and more recently a glycine-rich protein (GRPs) class. The HGRPs termed extensions have a characteristic S-P₄ repeat structure, the proline residues being hydroxylated and subsequently glycosylated.

A wide range of proline-rich proteins have now been isolated from dicotyledonous plants. Many of these have been shown to have cell wall associations although the functions remain as yet undefined. It is likely that in different cell types and in different species the PRPs present will vary.

The isolation of genomic and cDNA clones encoding PRPs has shown that this is a class of proteins containing some structurally more diverse sequences. It has been found that some of the genes encoding the PRPs display a highly regulated pattern of expression at the messenger RNA level and are induced by wounding, fungal infection, auxin or during nodulation. More recently two reports have come out showing that the expression of specific PRP genes is correlated with specific developmental events.

In contrast to the position for dicot plants, very little data has been accumulated in relation to PRPs from monocot species. Purification of an extensin-like protein from maize was reported and only low levels of hydroxyproline-residues were found. Subsequent to this a highly repetitive PRP cDNA sequence from maize has been

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published and mRNA expression studies have shown that highest levels are present in root tip and coleoptile. This protein shares a significant level of similarity with PRPs from dicot species with the exception that it contains a high proportion of threonine residues.

We have now identified a promoter sequence for each of two proline-rich protein (PRP) genes. Accordingly, the present invention provides a promoter having the nucleotide sequence:

- 10 (a) from -1369 to -49 upstream of the PRP140 gene,
or
(b) from -2316 to -12 upstream of the PRP378 gene;
the sequence optionally being modified by one or more base
substitutions, insertions and/or deletions and/or by an
15 extension at either or each end provided that the
thus-modified sequence is capable of acting as a promoter.

The invention also provides a DNA fragment comprising such a promoter operably linked to a heterologous gene encoding a protein. Additionally provided
20 is a vector which comprises a heterologous gene encoding a protein under the control of a promoter as above such that the gene is capable of being expressed in a plant cell transformed with the vector. A suitable vector is one in which the promoter is fused directly to the 5'-end of the
25 gene. The vector may further contain a region which enables the gene and the promoter to be transferred to and stably integrated in a plant cell genome. The vector is generally a plasmid.

Plant cells can be transformed with such a vector.
30 The invention therefore further provides plant cells which harbour a promoter as above operably linked to a heterologous gene encoding a protein. Transgenic plants may be regenerated from such plant cells. A transgenic plant can be obtained which harbours in its cells a promoter as
35 above operably linked to a heterologous gene encoding a protein. Seed may be obtained from the transgenic plants.

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The invention further provides a method of producing a desired protein in a plant cell, which method comprises:

(i) transforming a plant cell with a vector
5 according to the invention, the protein encoded by the gene under the control of the said promoter being the desired protein; and

(ii) culturing the transformed plant cell under conditions which allow expression of the protein.

10 The invention additionally provides a method of producing a transgenic plant capable of producing a desired protein, which method comprises:

(i) transforming a plant cell with a vector according to the invention, the protein encoded by the gene
15 under the control of the said promoter being the desired protein; and

(ii) regenerating plants from the transformed cells.

The desired proteins can be isolated from the
20 transformed plant cells obtained by the first method and from the plants obtained by the second method.

The present promoter is composed of the sequence upstream of the wheat PRP140 gene from base -1369 to base -49 or of the wheat PRP378 gene from base -2316 to base -12,
25 base 1 being A of the ATG translational start codon for PRP. The sequence of the PRP140 promoter can be deduced from Figure 6. The sequence of the PRP378 promoter can be deduced from Figure 5.

The promoter may be obtained by preparing a genomic
30 library of wheat DNA, screening the library for the PRP140 or PRP378 gene and digesting the sequence upstream of the wheat PRP140 or PRP378 gene with appropriate restriction enzymes and/or exonucleases. There is a NspBII restriction site at base -49 and a XbaI restriction site at base -1369
35 of the upstream sequence of the PRP140 gene. There is a NotI restriction site at base +72 and a HindIII site at base

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-2316 of the sequenc of the PRP378 gene.

Several plasmid vectors have been prepared which contain an upstream sequence of the wheat PRP140 or PRP378 gene comprising the promoter sequence. These vectors
5 include pIPKH-12/2.3, pXNot/1.45, pBIXN/1.3 and pHN/2.4. E. coli MC 1022 harbouring these vectors were deposited at the National Collection of Industrial and Marine Bacteria, Aberdeen, GB on 9 November 1989 under accession numbers NCIMB 40223, NCIMB 40224, NCIMB 40225 and NCIMB 40226
10 respectively. pBIXN/1.3 was deposited under the designation pIPXN/1.3.

The PRP140 promoter may be released from pXNot/1.45 by digesting the plasmid with XbaI and NspBII. The PRP378 promoter may be released from pHN/2.4 by linearisation with
15 NotI-BstXI, incubation with a nuclease, religation of the plasmid population, selecting a plasmid with a deletion at its 3'-end to base-12 of the PRP 378 gene and digesting the plasmid with HindIII.

The promoter sequence may be modified by one or
20 more base substitutions, insertions and/or deletions and/or by an extension at either or both ends. However, the modified promoter sequence must still be capable of acting as a promoter. Sequences from base -769 to base -49 of the upstream sequence of the wheat PRP140 gene, and shorter,
25 have been found not to be sufficient to direct expression of a protein at satisfactory levels. On the other hand, the sequence from base -961 to base -49 upstream of the wheat PRP140 gene does direct protein expression at a satisfactory level. Typically there is a degree of homology of at least
30 60% between a modified sequence and the unmodified natural sequence from base -1369 to base -49 upstream of the wheat PRP140 gene or from base -2316 to -12 upstream of the wheat PRP378 gene. The degree of homology may be at least 75%, at least 85% or at least 95%.

35 A longer sequence may be provided which extends upstream of base -1369 of the PRP140 gene or of base -2316

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of the PRP378 gene, for example to another restriction site. An extension upstream typically comprises the natural nucleotide sequence upstream of base -1369 of the PRP140 gene or of base -2316 of the PRP378 gene. Such a longer
5 sequence may be obtained from a genomic library of wheat DNA as above.

The upstream sequence of the wheat PRP 140 gene from base -961 to base -49 may also be modified by one or more base substitutions, insertions and/or deletions and/or
10 by an extension at either or both ends. Again, such a modified sequence must be capable of acting as a promoter. There may be a degree of homology of at least 60%, for example at least 75%, at least 85% or at least 95%, between the unmodified and modified longer sequences.

15 A modified promoter sequence may be obtained by introducing changes into the natural promoter sequence. This may be achieved by any appropriate technique, including restriction of the natural sequence with an endonuclease, insertion of linkers, use of an exonuclease and/or a
20 polymerase and site-directed mutagenesis. A shorter DNA sequence therefore may be obtained by removing nucleotides from the 5'-terminus or the 3'-terminus of the natural promoter sequence, for example using an exonuclease such as exonuclease III or BAL 31.

25 Whether a modified sequence is capable of acting as a promoter may be readily ascertained. The modified sequence is placed upstream of a protein coding sequence, such as the bacterial reporter gene β -glucuronidase as in the Example. Tobacco leaf discs can then be transformed.
30 The protein expressed by the transformed cells are then assayed, in the case of β -glucuronidase as described in Example 1.

The promoter may be operably linked to a heterologous gene encoding a protein. The heterologous gene
35 may encode any protein it is desired to express. By "heterologous" is that the gene is not naturally operably

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linked to the promoter. The gene does not therefore encode PRP140 in the case where the promoter is derived from the upstream sequence of the PRP140 gene or PRP378 in the case where the promoter is derived from the upstream sequence of the PRP378 gene. The protein may additionally comprise a transit peptide sequence at its N-terminus, encoded within the heterologous gene sequence.

The promoter is typically used to control the expression of genes in plant tissues. The protein whose expression is controlled by the promoter may be a protein encoded by a herbicide-resistance gene or a protein conferring biological control of pests or pathogens. The protein may therefore be an insecticidal protein, such as B. thuringiensis toxin, to give resistance to leaf-eating insects. Other uses to which the promoter may be put are the production of viral coat proteins to protect against viral infection, the production of high value proteins such as pharmaceuticals and the production of proteins to alter taste or nutritive value of forage grasses, etc.

The promoter sequence may be fused directly to a heterologous gene or via a linker. The linker sequence may comprise an intron. Excluding the length of any intron sequence, the linker may be composed of up to 45 bases, for example up to 30 or up to 15 bases. The linker sequence may comprise a sequence encoding a transit amino acid sequence, for example a transit sequence capable of directing a protein to a chosen subcellular locality such as the chloroplasts or mitochondria. The linker sequence may comprise a sequence having enhancer characteristics, to boost expression levels.

DNA fragments and vectors can be prepared in which the promoter is operably linked to a heterologous gene. The fragments and vectors may be single or double stranded. Plant cells can be transformed by way of such fragment directly or by way of such a vector. The vector incorporates the heterologous gene under the control of the

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promoter. The vector contains regulatory elements capable of enabling the gene to be expressed in a plant cell transformed with the vector. Such regulatory elements include, besides the promoter, translational initiation and/or termination sequences. The vector typically contains too a region which enables the chimaeric gene and associated regulatory control elements to be transferred to and stably integrated in the plant cell genome.

The vector is therefore typically provided with transcriptional regulatory sequences and/or, if not present at the 3'-end of the coding sequence of the gene, a stop codon. A DNA fragment may therefore also incorporate a terminator sequence and other sequences which are capable of enabling the gene to be expressed in plant cells. An enhancer or other element able to increase or decrease levels of expression obtained in particular parts of a plant or under certain conditions may be provided in the DNA fragment and/or vector. The vector is also typically provided with an antibiotic resistance gene which confers resistance on transformed plant cells, allowing transformed cells, tissues and plants to be selected by growth on appropriate media containing the antibiotic.

Transformed cells are selected by growth in an appropriate medium. Plant tissue can therefore be obtained comprising a plant cell which harbours the heterologous gene under the control of the promoter, for example in the plant cell genome. The gene is therefore expressible in the plant cell. Plants can then be regenerated which include the heterologous gene and the promoter in their cells, for example integrated in the plant cell genome, such that the gene can be expressed. The regenerated plants can be reproduced and, for example, seed obtained.

A preferred way of transforming a plant cell is to use Agrobacterium tumefaciens containing a vector comprising the promoter operably linked to the heterologous gene. A hybrid plasmid vector may therefore be employed which

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comprises:

(a) the heterologous gene under the control of the promoter and other regulatory elements capable of enabling the gene to be expressed when integrated in the genome of a plant cell;

(b) at least one DNA sequence which delineates the DNA to be integrated into the plant genome; and

(c) a DNA sequence which enables this DNA to be transferred to the plant genome.

Typically the DNA to be integrated into the plant cell genome is delineated by the T-DNA border sequences of a Ti-plasmid. If only one border sequence is present, it is preferably the right border sequence. The DNA sequence which enables the DNA to be transferred to the plant cell genome is generally the virulence (vir) region of a Ti-plasmid.

The heterologous gene and its transcriptional and translational control elements, including the promoter, can therefore be provided between the T-DNA borders of a Ti-plasmid. The plasmid may be a disarmed Ti-plasmid from which the genes for tumorigenicity have been deleted. The gene and its transcriptional and control elements, including the promoter, can, however, be provided between T-DNA borders in a binary vector in trans with a Ti-plasmid with a vir region. Such a binary vector therefore comprises:

(a) the heterologous gene under the control of the promoter and other regulatory elements capable of enabling the gene to be expressed when integrated in the genome of a plant cell; and

(b) at least one DNA sequence which delineates the DNA to be integrated into the plant genome.

Agrobacterium tumefaciens, therefore, containing a hybrid plasmid vector or a binary vector in trans with a Ti-plasmid possessing a vir region can be used to transform plant cells. Tissue explants such as stems or leaf discs may be inoculated with the bacterium. Alternatively, the

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bacterium may be co-cultured with r generating plant protoplasts. Plant protoplasts may also be transformed by direct introduction of DNA fragments which encode the heterologous gene and in which the promoter and appropriate
5 other transcriptional and translational control elements are present or of a vector incorporating such a fragment. Direct introduction may be achieved using electroporation or polyethylene glycol, microinjection or particle bombardment.

Plant cells from monocotyledonous or dicotyledonous
10 plants can be transformed according to the present invention. Monocotyledonous species include barley, wheat, maize and rice. Dicotyledonous species include tobacco, tomato, sunflower, petunia, cotton, sugarbeet, potato, lettuce, melon, soybean, canola (rapeseed) and poplars.
15 Tissue cultures of transformed plant cells are propagated to regenerate differentiated transformed whole plants. The transformed plant cells may be cultured on a suitable medium, preferably a selectable growth medium. Plants may be regenerated from the resulting callus. Transgenic plants
20 are thereby obtained whose cells harbour the promoter operably linked to the heterologous gene, for example integrated in their genome. The gene is consequently expressible in the cells. Seed from the regenerated plants can be collected for future use.

25 The following Examples illustrate the present invention. In the accompanying drawings:

Figure 1 shows the restriction map of the insert DNA of phage 3cX2;

Figure 2 shows the restriction map of the insert
30 DNA of phage E3.2;

Figure 3 shows the sequence of the PRP 378 gene;

Figure 4 shows the sequence of the PRP 140 gene;

Figure 5 shows the regulatory sequence upstream of the PRP 378 gene;

35 Figure 6 shows the regulatory sequence upstream of the PRP 140 gene;

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Figure 7 shows the construction of pBIHN/2.4;

Figure 8 shows the construction of pBIXN/1.3;

Figure 9 shows the construction of pBID -234, -400, -540, -769 and -961 N;

5 Figure 10 shows the construction of pIPKH -6, -12 and -50/2.3;

Figure 11 shows the construction of pIPH-6/2.3;

Figure 12 shows the construction of pIP S-6/762 and pIP P-6/615;

10 Figure 13 shows the construction of pIPD-6/1839, 1510, 1289, 1023 and 816;

Figure 14 shows the sequences of the fusion junctions of constructs based on BIN 19;

Figure 15 shows the sequences of the fusion
15 junctions of constructs based on pUC19;

Figure 16 shows the results of Gus assays for the PRP140 promoter deletions;

Figure 17 shows the results of Gus assays for the PRP 378 deletions; and

20 Figure 18 shows the nucleotide sequence of the cDNA clone (WPRP1).

EXAMPLE 1

1. Isolation of PRP Genes

The strategy employed to isolate wheat PRP
25 promoters involved first constructing a genomic library of wheat DNA and then screening of this library using a cDNA probe for a PRP gene.

High molecular weight DNA was isolated from dark grown shoots of Triticum aestivum cv. Chinese Spring
30 (Lazarus et al, Plant Mol. Biol. 5, 8-24, 1985). Conditions for partial digestion with Sau3A were established. DNA fragments of 18-25 kb were purified by size fractionation on sucrose density gradients. Lambda Charon 35 (Loenen and Blattner, Gene 26, 171-179, 1983) vector DNA was prepared by
35 digestion with BamHI and purification on sucrose density gradients. Vector DNA (1.5 µg) and wheat DNA (3.5 µg) were

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ligated at high concentration (500 ng/ μ l) using T4 DNA ligase and subsequently packaged in vitro using commercially available extracts (Stratagene).

About 2.6×10^6 recombinants were plated onto the 5 host E. coli K803 and DNA lifts from the plaques taken onto nitrocellulose filters. The wheat PRP cDNA WPRP1 (Reference Example) was random primer labelled using 32 P dATP and used to probe the library lifts. Positively hybridizing plaques were purified to homogeneity by several rounds of screening 10 and then DNA from these positive phage was purified by further analysis.

2. Characterization of the PRP Genes

Restriction maps were made of the insert DNA of the positive phages 3cX2 (Fig. 1) and E3.2 (Fig. 2) and the 15 position and orientation of the PRP genes determined by hybridization studies. DNA fragments covering these regions were subcloned into the plasmid vector pUBS1, which is a pUC19 derivative containing the polylinker region of the Bluescript plasmid of Stratagene (Raines et al, Nucl. Acids. 20 Res. 16, 7931-7942, 1988). Overlapping sequence was obtained from these clones by Exonuclease III digestion and double-stranded dideoxy nucleotide sequence analysis. Data were assembled and analysed using the Staden packages. The extent of the sequence determined is indicated on Figs 1 and 25 2.

DNA sequence comparisons between the PRP 378 gene (Fig. 3) and the cDNA probe used to isolate it revealed a few base differences which suggested that the gene copy isolated may represent an allele of the cloned mRNA. The 30 PRP 140 gene (Fig. 4) is evidently a novel form of PRP.

3. PRP Upstream Sequences

The regulatory sequence upstream of the PRP 378 and PRP 140 genes, including all of those used in the constructs described below, are given in Figs 5 and 6 respectively. 35 Restriction enzyme sites used during subsequent cloning procedures are indicated and underlined. It was proposed

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that these sequences would contain all of the promoter and other regulatory elements necessary to direct correct expression of protein coding sequences placed downstream under their control.

5 In order to test this proposal and assess the usefulness of the PRP 378 and PRP 140 promoters in the context of plant transformation a series of constructs were prepared. In each case part of the PRP 378 or PRP 140 upstream regions was placed in front of the bacterial
10 reporter gene β -glucuronidase and the nopaline synthase terminator sequences in a suitable vector. For the PRP 378 promoter this was a high copy number pUC19 based plasmid for transient expression in Zea mays protoplasts. For the PRP 140 promoter this was a BIN19 based vector suitable for
15 transformation into Nicotiana tabacum. In both cases this allows expression, controlled by the PRP promoters, to be detected by an enzyme assay for β -glucuronidase (Gus) activity using a fluorogenic substrate in extracts from transformed plants or protoplasts.

20 (a) pIPPKGT and pBIPKGT

pIPPKGT is the EcoRI/SalI insert of pRAJ275 (Jefferson, Plant. Mol. Biol. Reporter 5, 387-405, 1987) blunted with Klenow fragment of E. coli DNA polymerase I, and ligated into the Klenow blunted Asp718 site of
25 pUCPolyTer (the nopaline synthetase terminator fragment cloned as a SstI/EcoRI fragment into pUC19). The corresponding BIN19 based plasmid is pBIPKGT. pIPPKGT and pBIPKGT can be obtained from each other.

(b) BIN19 Based Constructs (pBI series)

30 (i) Construction of the plasmid pBIHN/2.4 containing 2318 bp of sequence 5' of the PRP 378 translation initiation codon and 24 amino acids of PRP 378 coding sequence in a translational fusion to β -glucuronidase (Fig. 7)

A 2.4 Kb HindIII-NotI fragment of the PRP 378 gene
35 (Fig. 1) was subcloned into the plasmid vector pUBS1 to form the plasmid pHN/2.4. pHN/2.4 was linearised with NotI and

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the site filled in using Klenow fragment of E. coli DNA polymerase I. The insert was excised by digestion with HindIII and inserted into HindIII-SmaI digested pBI101.3 (Jefferson, Plant. Mol. Biol. Reporter 5, 387-405, 1987) to form pBIHN/2.4. The fusion junction was checked by nucleotide sequencing and its sequence along with the predicted amino acid sequence is shown in Fig. 14.

(ii) Construction of the plasmid pBIXN/1.3, a transcriptional fusion of 1321 bp of 5' non-coding sequence from PRP 140 to β -glucuronidase (Fig. 8)

A 1.45 kb XbaI-NotI fragment of the PRP 140 gene (Fig. 2) was subcloned into the plasmid vector pUBS1 to form pXNot/1.45. A 1.3 kb fragment was excised by digestion with XbaI and NspBII (Fig. 8) and inserted into XbaI-SmaI digested pBIPKGT to form pBIXN/1.3. The sequence of the fusion junction is shown in Fig. 14.

(iii) Construction of the plasmids pBID -234, -400, -540, -769 and -961 N representing deletion of the PRP 140 promoter from its 5' end (Fig. 9)

The plasmid pXNot/1.45 (Fig. 8) was linearised by digestion with XbaI-PstI. Incubation with Exonuclease III for various periods resulted in deletion of varied lengths of sequence from the XbaI end. Treatment with Nuclease SI and the Klenow fragment of E. coli DNA polymerase enabled the plasmid population to be religated as closed circles. Nucleotide sequencing was employed to determine the extent of deletion from the XbaI end of a representative portion of the population.

On the basis of the information provided by sequencing, plasmids with deletion to within 234, 400, 540, 769 and 961 nucleotides 5' of the translation initiation codon of PRP 140 were chosen for evaluation of promoter activity (Fig. 6). The 5 restriction fragments were excised by digestion with HindIII and NspBII and inserted into

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HindIII-SmaI digested pBIPKGT to form pBID -234, -400, -540, -769 and -961 N. In all 5 constructs the sequence of the fusion junction is identical to that of pBIXN/1.3 (Fig. 14).

(c) pUC19 Based Constructs (PIP series)

- 5 (i) Construction of the plasmids pIPKH -6, -12 and -50/2.3 representing deletion of the PRP 378 promoter from its 3' end (Fig. 10)

The plasmid pHN/2.4 (Fig. 7) was linearised by digestion with NotI-BstXI. Incubation with Exonuclease III
10 for various periods resulted in deletion of varied lengths of sequence from the NotI end. Treatment with Nuclease SI and the Klenow fragment of *E. coli* DNA polymerase enabled the plasmid population to be religated as closed circles. Nucleotide sequencing was employed to determine the extent
15 of deletion from the NotI end of a representative portion of the population.

On the basis of the information provided by sequencing, plasmids pHD -6, -12, -50/2.3 with inserts which extended approximately 2.3 kb downstream from the HindIII
20 sites to within 6, 12 and 50 nucleotides 5' of the translation initiation codon of PRP 378 were chosen for evaluation of promoter activity (Fig. 5). The 3 respective plasmids were linearised by a partial SstI digestion and the overhang blunted with Mung Bean Nuclease. The 3 inserts
25 were excised by digestion with HindIII and inserted into HindIII-SmaI digested pIPKGT to form pIPKH -6, -12, -50/2.3. The sequences of the fusion junctions are shown in Fig. 15.

(ii) Construction of pIPH-6/2.3 (Fig. 11)

- 30 This plasmid is similar to pIPKH-6/2.3 except that the full length PRP 378 promoter has been inserted into pBI201.1 (Jefferson, 1987) and has a different junction (Fig. 15) without a "Kozak" consensus initiating ATG.

- (iii) Construction of pIP S-6/762, P-6/615 representing
35 short forms of the PRP 378 promoter deleted from its 5' end

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(Fig. 12)

The plasmid pIPH-6/2.3 (Fig. 11) was linearised by PstI or partial SstI digestion and the overhang blunted with Mung Bean Nuclease. Inserts with respectively 762 and 615 5 nucleotides of the PRP 378 promoter in a cassette with the GUS gene and the NOS terminator were excised by digestion with EcoRI and inserted into EcoRI/SmaI digested pUC19. In both constructs the sequence of the fusion junction is identical to that of pIPH-6/2.3 (Fig. 15).

- 10 (iv) Construction of pIPD-6/1839, 1510, 1289, 1023, 816 representing longer forms of the PRP 378 promoter deleted from its 5' end (Fig. 13)

The plasmid pHN/2.4 (Fig. 7) was linearised by digestion with HindIII-ApaI. Incubation with Exonuclease 15 III for various periods resulted in deletion of varied lengths of sequence from the HindIII end. Treatment with Nuclease SI and the Klenow fragment of E. coli DNA polymerase enabled the plasmid population to be religated as closed circles. Nucleotide sequencing was employed to 20 determine the extent of deletion from the HindIII end of a representative portion of the population.

On the basis of information provided by sequencing plasmids pD-1839, -1510, -1289, -1023 and -816 N with inserts extending from 1839 to 816 nucleotides 5' of the 25 translation initiation codon of PRP 378 were digested with KpnI-SstII. The excised fragments were ligated with EcoRI-partial SstII digested pIPH-6/2.3 (providing the proximal portion of the PRP 378 promoter in conjunction with the GUS gene and the NOS terminator) which in turn was 30 ligated into EcoRI-KpnI digested pUC19. Once again in all constructs the sequence of the fusion junction is identical to that of pIPH-6/2.3 (Fig. 15).

4. Introduction of PRP Promoter Fusions into Nicotiana

Constructs were mobilised from Escherichia coli 35 MC1022 into Agrobacterium tumefaciens LBA4404 as described (Bevan, Nucl. Acids. Res. 12, 8711-8721, 1984). Leaf discs

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f Nicotiana tabacum var. Samsun were transformed as described (Horsch et al, Science 223, 496-498, 1984) and selected on shooting medium containing 100 µg/ml kanamycin. Typically between 5 and 15 plants containing each construct 5 were regenerated and assayed.

5. β-Glucuronidase Assay of Transformed Plants

The activity of the PRP promoters in individual transformants was determined by measuring β-glucuronidase activity in leaf extracts. Tissue extracts were prepared 10 and analysed for fluorescence of the reaction product 4-methyl umbelliferone as described (Jefferson, 1987). Reactions were usually incubated at 37°C for 4 hours with aliquots sampled at 2 hour intervals. The protein concentration in each extract was measured to allow direct 15 comparisons to be made between them (using a Bio-Rad kit).

Results of assays on PRP140 promoter derived constructs expression in nmol 4-methyl umbelliferone produced/min/mg protein are shown in Fig. 16. Because of the variation commonly observed between individual 20 tranformed plants it is difficult to ascertain whether dissection of the PRP 140 promoter from 1369 to 961 nucleotides in length leads to a significant loss of activity. However, it is clear that the 769 and 540 nucleotide forms have substantially less activity and that 25 the 400 and 234 nucleotide forms are effectively inactive.

6. Introduction of PRP 378 Promoter Fusions into Protoplasts Derived from a Suspension Culture of Zea mays Anther Cells

The pUC19 based constructs were assayed for 30 transient expression by Ca⁺⁺- PEG mediated transformation of Zea mays anther derived cell suspension (cell line Ba Tan Bai) protoplasts based on the method of Chen et al. (Chen, D.F., Tassie, A., Dale, P.J., Goldsbrough, A.P., Liang, W., Bevan, M.W., Flav 11, R.B. and Xia, Z.A. (1988). In: Puite, 35 K.J., Dons, J.J.M., Huizing, H.J., Kool, A.J., Koornneef, M. and Krens, F.A. (eds) Progr ss in Plant Protoplast Research.

- 17 -

Kluwer Academic Publishers, Dordrecht). Typically for each construct 3 replicates of 50 μ g of DNA and 1×10^6 protoplasts were performed. As controls protoplasts were also transformed with identical amounts of herring sperm DNA and the plasmid CG20 carrying one copy of the 850 bp form of the cauliflower mosaic virus (CaMV) 35S promoter.

7. β -Glucuronidase Assay of PEG mediated Transformed Zea mays Protoplasts

Lysates of transformed protoplasts were assayed for β -glucuronidase in the same manner as that for transformed plants (Jefferson, 1987). Expressing activity per 1×10^4 protoplasts allowed comparisons to be made between constructs tested in the same experiment.

Results of assays on PRP 378 promoter derived constructs expressed in nmol 4-methyl umbelliferone produced/min/ 1×10^4 protoplasts are shown in Table 1 and Fig. 17.

Table 1. Gus Assays: PRP378 Promoter 3' Deletions

<u>Construct</u>	<u>nMOL of 4-MU/min/1×10^4 protoplasts</u>
CG20 (CaMV 35S)	3.59 ± 2.96 (100%)
Herring sperm DNA	0.0308 (0.858)
PIPKH -6 /2.3	9.65 ± 0.63 (269)
-12	11.92 ± 4.22 (332)
-50	4.73 ± 0.66 (132)

The 2.3 kbp form of the PRP 378 promoter is significantly stronger than the 850 bp form of the CaMV35S promoter in maize protoplasts (Table 1). In addition it is sensitive to relatively short deletions from its 3' end. Thus PIPKH-12/2.3, the construct that places the promoter in almost its native context (Fig. 15) with respect to the initiating "ATG" of the GUS gene, exhibits highest levels of expression; whilst activity declines if, in effect, the promoter is moved (Fig. 15) further or closer to the ATG.

Figure 17 illustrates the effect of deletions from

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5' end of the PRP 378 promoter. A deletion of 800 bp reduces activity to less than 60% of the full length form, and activity continues to decline with extent of deletion. Although the full length form of the PRP 378 promoter in this experiment does not have the same level of activity with respect to CaMV 35S as those in Table 1, its fusion junction is different and the initiating ATG of the β -glucuronidase gene does not have the "Kozak" consensus (Fig. 15).

10 REFERENCE EXAMPLE

A cDNA, designated WPRP1, encoding a wheat PRP has been isolated and sequenced. The WPRP1 clone was detected whilst a wheat cDNA library was being screened using antibodies raised against chloroplastic fructose-1,5-bisphosphatase. Why this antibody should cross-react with the WPRP1 clone is not known. The cDNA isolated initially was not full length (1.3 Kb). The complete sequence came from a longer cDNA obtained by rescreening the cDNA library using the shorter clone.

20 The complete nucleotide sequence of the cDNA clone (WPRP1) from wheat with its deduced amino acid sequence is shown in Figure 18. This 1548 bp cDNA contains the complete coding sequence of 1137 base pairs with 101 bp of 5' non-coding sequence and 310 bp of 3' non-coding sequence

25 terminating in a poly-adenylation tail.

EXAMPLE 2

The localisation of Gus expression in tissues of the transformed tobacco plants of Example 1.4 was investigated. Assays were conducted as described in Example 1.5 except that tissues other than leaf were also assayed. The results are shown in Table 2 below. The values shown are pmol/min/ug protein. The numbers in brackets refer to ranking of tissue expression level within each individual transformed plant. There are significant

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levels of expression in all tissues. The levels of expression in petioles and stems compare favourably with CaMV 35S.

Table 2

5		<u>leaf</u>	<u>petiole</u>	<u>root</u>	<u>stem</u>
PRP140: 961	3	7.93 (5)	102.75 (1)	18.82 (3)	16.96 (4)
	4	10.38 (3)	170.16 (1)	5.29 (4)	63.87 (2)
	6	6.28 (4)	65.02 (1)	5.65 (5)	30.53 (2)
1369	2	17.22 (3)	95.45 (1)	3.29 (5)	51.86 (2)
10	5	4.58 (3)	104.64 (1)	2.04 (5)	24.52 (2)
	8	16.60 (3)	305.35 (1)	7.32 (5)	40.87 (2)
	12	6.64 (5)	76.25 (1)	6.65 (4)	33.20 (2)
CaMV		39.96 (4)	58.25 (2)	50.14 (3)	62.43 (1)
BIN(0)		0.35	1.31	3.62	0.67
15		<u>apical</u>			
PRP140: 961	3	19.55 (2)			
	4	1.96 (5)			
	6	9.08 (3)			
1369	2	17.21 (4)			
20	5	4.31 (4)			
	8	26.70 (4)			
	12	12.41 (3)			
CaMV		32.48 (5)			
BIN(0)		0.32			

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CLAIMS

1. A promoter having the nucleotide sequence:
 - (a) from -1369 to -49 upstream of the PRP140 gene, or
 - (b) from -2316 to -12 upstream of the PRP 378 gene;
- 5 the sequence optionally being modified by one or more base substitutions, insertions and/or deletions and/or by an extension at either or each end provided that the thus-modified sequence is capable of acting as a promoter.
2. A promoter according to claim 1, having a
- 10 sequence from -961 to -49 upstream of the PRP140 gene, optionally modified as specified in claim 1.
3. A DNA fragment comprising a promoter as claimed in claim 1 or 2 operably linked to a heterologous gene encoding a protein.
- 15 4. A vector which comprises a heterologous gene, encoding a protein, under the control of a promoter as claimed in claim 1 or 2, such that the gene is capable of being expressed in a plant cell transformed with the vector.
5. A vector according to claim 4, wherein the
- 20 promoter is fused directly to the 5'-end of the said gene.
6. A vector according to claim 4 or 5, which further contains a region which enables the gene and the promoter to be transferred to and stably integrated in a plant cell genome.
- 25 7. A vector according to any one of claims 4 to 6, which is a plasmid.
8. A plant cell which has been transformed with a vector as claimed in any one of claims 4 to 7.
9. A plant cell which harbours a promoter as
- 30 claimed in claim 1 or 2 operably linked to a heterologous gene encoding a protein.
10. A transgenic plant which has been regenerated from plant cells as claim d in claim 8 r 9.
11. A transgenic plant which harbours in its cells
- 35 a promot r as claim d in claim 1 or 2 operably linked to a heterologous gene encoding a protein.

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12. Seed obtained from a transgenic plant as claimed in claim 10 or 11.

13. A method of producing a desired protein in a plant cell, which method comprises:

- 5 (i) transforming a plant cell with a vector as claimed in any one of claims 4 to 7, the protein encoded by the gene under the control of the said promoter being the desired protein; and
- (ii) culturing the transformed plant cell under
10 conditions which allow expression of the protein.
14. A method of producing a transgenic plant capable of producing a desired protein, which method comprises:
- 15 (i) transforming a plant cell with a vector as claimed in any one of claims 4 to 7, the protein encoded by the gene under the control of the said promoter being the desired protein; and
- (ii) regenerating plants from the transformed cells.

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Fig. 1.

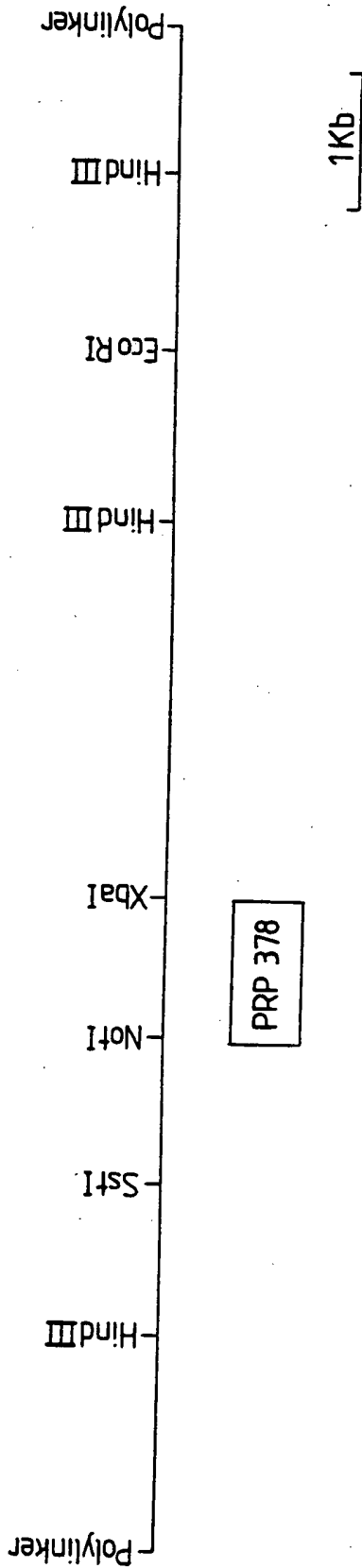
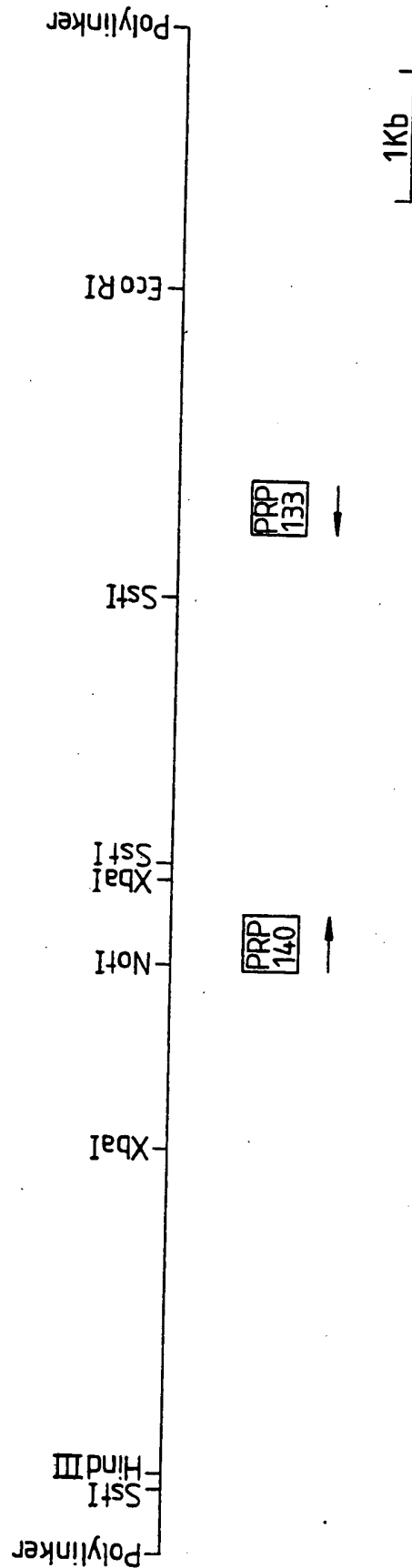


Fig. 2.



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Fig. 3.

2319 ATGGCGAGGCACAGCCTCCTTGCCGTGCTCCTCGTGGGCTAGTCGGGCTCCGGCTTC 2378
 M A R H S L L A V L L V G L V A A S G F
 2379 AGCCAGGCGCGCGCTGGCCGTGGCTCGCTGAGAAGCTCCGGAGCGGAGCCCAAG 2438
 S Q A A A A G R G L A E K L P E P E P K
 2439 CCGACGCCGTACCCGGAGCCCAAGCCGGAACCCCAAGCCAGAGCCCAATGCCTAAGCCTGAA 2498
 P T P Y P E P K P E P K P E P M P K P E
 2499 CCCATGCCAAAGCCAGAGCCTTAAGCCTGAACCCATGCCAAAGCCAGAGCCTAAGCCTATG 2558
 P M P K P E P K P E P M P K P E P K P M
 2559 CCCAAACCTGAGCCCATGCCAAAGCCAGAGCCCAACCCGGAACCGAAGCCGAGCCGATG 2618
 P K P E P M P K P E P K P E P K P E P M
 2619 CCAAAGCCCGAACCAAAACCTGAGCCCAAGCCTTACCCGATGCCCAACCTGAGCCTAAG 2678
 P K P E P K P E P K P Y P M P K P E P K
 2679 CCTGAGCCCAAGCCCGAGCCTATGCCAAACCTGAACCAAGCCAGAGCCCAAGCCCGAG 2738
 P E P K P E P M P K P E P K P E P K P E
 2739 CCTATGCCAAACCTGAACCAAGCCCGAGCCCAAGCCTGAGCCTATGCCAAACCTGAG 2798
 P M P K P E P K P E P K P E P M P K P E
 2799 CCTAAGCCGATGCCAAACCCGAACCGAAGCCAGAGCCCAAGCCTGAGCCGATGCCTAAG 2858
 P K P M P K P E P K P E P K P E P M P K
 2859 CCAGAACCGAAGCCAGAGCCCAAGCCTGAGCCTATGCCAAACCTGAACCAAGCCAGAG 2918
 P E P K P E P K P E P M P K P E P K P E
 2919 CCCAAACTCGAGCCGATGAAGCCCGAGCCTAAGCCGTGCCAAACCCGAACCTAAGCCA 2978
 P K L E P M K P E P K P V P K P E P K P
 2979 GATCCCAAGCCTGAGCCGATGCCAAACCGGAGCCCAAGCCAGAGCCCAAGCCTGAGCCG 3038
 D P K P E P M P K P E P K P E P K P E P

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Fig. 3(cont.)

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3039  ATGCCAAACCCGAACCGAGCCGGAGCCCAAGCCTTACCCAATGCCCAAACTGAGCCT  3098
      M P K P E P K P E P K P E P K P Y P M P K P E P
3099  AAGCCTGAGCCCAAGCCTGAGCCGATGCCCAAAACCAAGCAACCAAGCCAGAGCCCAACCC  3158
      K P E P K P E P M P K P E P K P E P K P E P K P
3159  GAGCCGATGAAGCCAGAGCCTAAGCCAAATGCCAAAGCCGAACCGAAGCCAGAGCCCAAG  3218
      E P M K P E P K P M P K P E P K P E P K P E P K
3219  CCTGAGCCGATGCCCAAGCCCGAGCCCAAGCCTATGCCGAAGCCAGAGCCCAACCCAGAG  3278
      P E P M P K P E P K P E P K P M P K P E P K P E
3279  CCGATGCCCAAGCCAGAGCCTAAGCCCGAACCATTGCCCTAAACCAAGAGCCTAAGCCCGAA  3338
      P M P K P E P K P E P L P K P E P K P E P K P E
3339  CCAATGCCCTAAACCAAGAGCCCAAGCCTGAACCTATGCCCTAAGCCGGAGCCCAAGCCTGAA  3398
      P M P K P E P K P E P M P K P E P K P E P K P E
3399  CCCGAACCAAGCCAGAGCCACCTCCGAAGGCAAGCCGCAATGACTGACAATTGATGT  3458
      P E P K P E P P P K G K P P M T D N *
3459  GATACTCACATATGACAGCTGAAGGAGGAGATCGACCCCGTCCGGAGCCACGGTGTGCTG  3518
      XbaI
3519  TTTCCTAGA 3526

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Fig. 4.

10 30 50
 1 ATGGCGAGGCATCGCCTCCTTGCCGTGCTCATGCTCCTCGTGGGGTAGTGGCAGCCTCC 60
 M A R H R L L A V L M L L V G V A A S
 70 90 110
 61 ACTTTCCACCAAGCGCGCGCGTGGCGGGCCTTGCTGCCGTCGAGAAAGTTTGGGGAC 120
 T F H Q A A A A G R G L A A V E K F A D
 130 150 170
 121 CTGGAGCCAAAGCCAAACCTAAACAAGGCAATGCCCAACCAATGCCGAAGCCTGAA 180
 L E P K P K P K Q E A M P K P M P K P E
 190 210 230
 181 CCCAAGCCGATGCCAAAGCCCGAGCCCAAGCCCAAGCCGATGCCTAAACCTGAA 240
 P K P M P K P E P K P K P K P M P K P E
 250 270 290
 241 CCCAAGCCAGAGCCCAAGCCCAAGCCGATGCCCAACCTGAACCAAGCCAGAGCCCAAG 300
 P K P E P K P K P M P K P E P K P E P K
 310 330 350
 301 CATAAACCAATGCCTAAGCCAGAGCCTAAGCCGGAACCTATGCCCTAAGCCGAGCCCAAG 360
 H K P M P K P E P K P E P M P K P E P K
 370 390 410
 361 CCTAAACCTGACCCGAAACTAGAGCCACCACCAAGCACAAAGCCGCCAACAGCTTACAAT 420
 P K P D P K L E P P P K H K P P T A Y N
 430 450 470
 421 TGATGGGATACTATATATGACAACAGAAAGATCAAGGAGATCATGGCCGGGCCACAATC 480
 *
 490 510 530
 481 GCGATATTCTGGAATAAGTAGTAGTAACAATGTCAATTCATCCGGCTAGCTCGATAG 540

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Fig. 4 (cont.)

541	550	CGATGCATCTTTCTGTATTCTATGTAATTTTCGTTACATTTATAAAGTTTCTGTCCATGCA	570	590	600
	610		630	650	
601	670	TCCATGATGAGTATTACTACTTTTGTAAATTGTTATTGCTTGTGGTGTAAAGTTCAATTTT	690	710	660
661		GTACGTGACTCGTTGTTCTATTGTTGTATCGCATGAATGAGTTATTTTGTCTAGA		.XbaI	715

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Fig. 5.

[illegible]

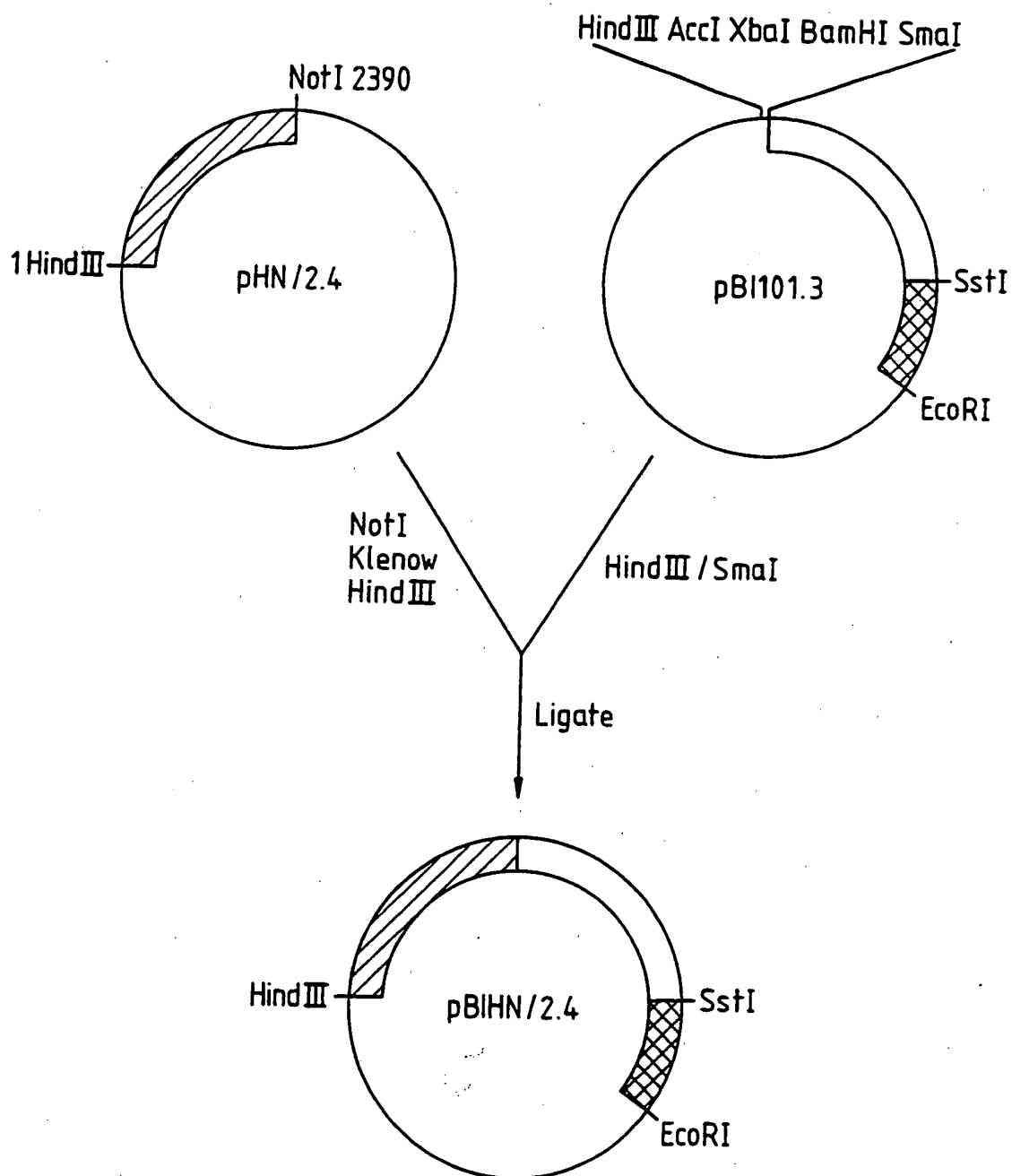
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Fig. 6.

XbaI
 1 TTCTAGAAAGTACAAAACTACCAATTCCTCTAGTGTGTCGAAATATTCCTAAATCTCTTAGGTTTCTTTTAGAAAAAGTATATGCTTCTACTAGAAATTA
 101 AAGGAATAATGTCGATATCAAGATGATATCAATTTACACCCGGCCCTCTGCGAAAAACACGGTGTCAAAAGTTATTTCAAGACATCGAGCTGCGACAACTTAAT
 201 TATTTTAAACAAAAATAAAAAAAGAACGATAATAACACGGCGAGCGACCAACATCGAGCGCAGTGCAGAGAAGCGEACGAGGGGTGTGAGCCCTTATATCATTA
 301 TCATGCCATTCACCTAACTCCCTTTGNTAOCAGCAAGAAATTTGTGTTGTTACTTCTCTCAANTGCGCCAAOCGACANTCTATGATCGAAGTAAGAGA
 401 ATGCTTCTCTGATTTTCTCTATATTTGTACAGTGTACAACCTCTCTGTTTATGAGTATTTGGGGTTTTCTCTCTTTAGTGAATGTAAACCAAGAAAG
 501 AAGGCAGCGCTGCTGCAATTTTGGCTTAAATTCATGATATTTGGGGTTGGGTGGTCAATGCTAGGCTTAAACCGCCAAAGTTATTTAACTTTCAGCGCATCTTA
 601 CCTATCCATATTTCTAOGTTGATGAGGTACGTAGAAAAATAGTAAAGGAGAGATGAAGGTTGACACATGTTAGTACAAAGAAAGCGTCAGCGAGTCCAAAC
 701 CATGCATGCTAATCAACCGGGAACATAOGCCGGGCTGGAAOCCATGCCCTACTCTCTCAANTGAATAAATATAGTGTAGGCGGAGTGAAGTGTAGAAAAAGG
 801 CCATGGTGCATGCTGCTGTGGGAAACGGATGGGGGGTCAATTTTGTGTGACCAACCTGCTGTGTCGACAGAGGATACCCGTAGCTTACAGCTGGCCCTC
 901 TCTACTTACGTTAAACCACTACTATCTTTTATCATATGTTATGTTTGTATGTCATGTCGCTCTGTACCTTACCGGCAACTGCTCAGCAOCCACACAAOCCATAC
 1000 AATAACAGATCACATCTATATATATACATCTTAAAGTATATATATGGTGCCCTACATAGAGTGTGTGTCGTTGAGCAAGGCAACTAAACAGCATTT
 1100 TCCTGTTTGCAGACGGGTGCAACTAATAGAGTTGAAATACTATTATCTGGATAATGTTGTTGTCGCGANTCCACACCCACCGAAGCTGGTTTACAGGGA
 1200 TTGTGGCACACGGTCTGNTTGGCTCTCTCCACCCCTATTAATAAACACACCGTATGTGTCTCCACTTCTCCACAGAGCACACCCATCCACCATCC
 1300 ATCCATCCAGGCTAGACAGCGGGGGAACAACAAGTGGGCGCCCTAGCTGCGAGCAACAGGGCGAGCGCAATG
 NspBII
 1301 ATCCATCCAGGCTAGACAGCGGGGGAACAACAAGTGGGCGCCCTAGCTGCGAGCAACAGGGCGAGCGCAATG

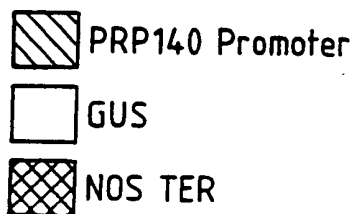
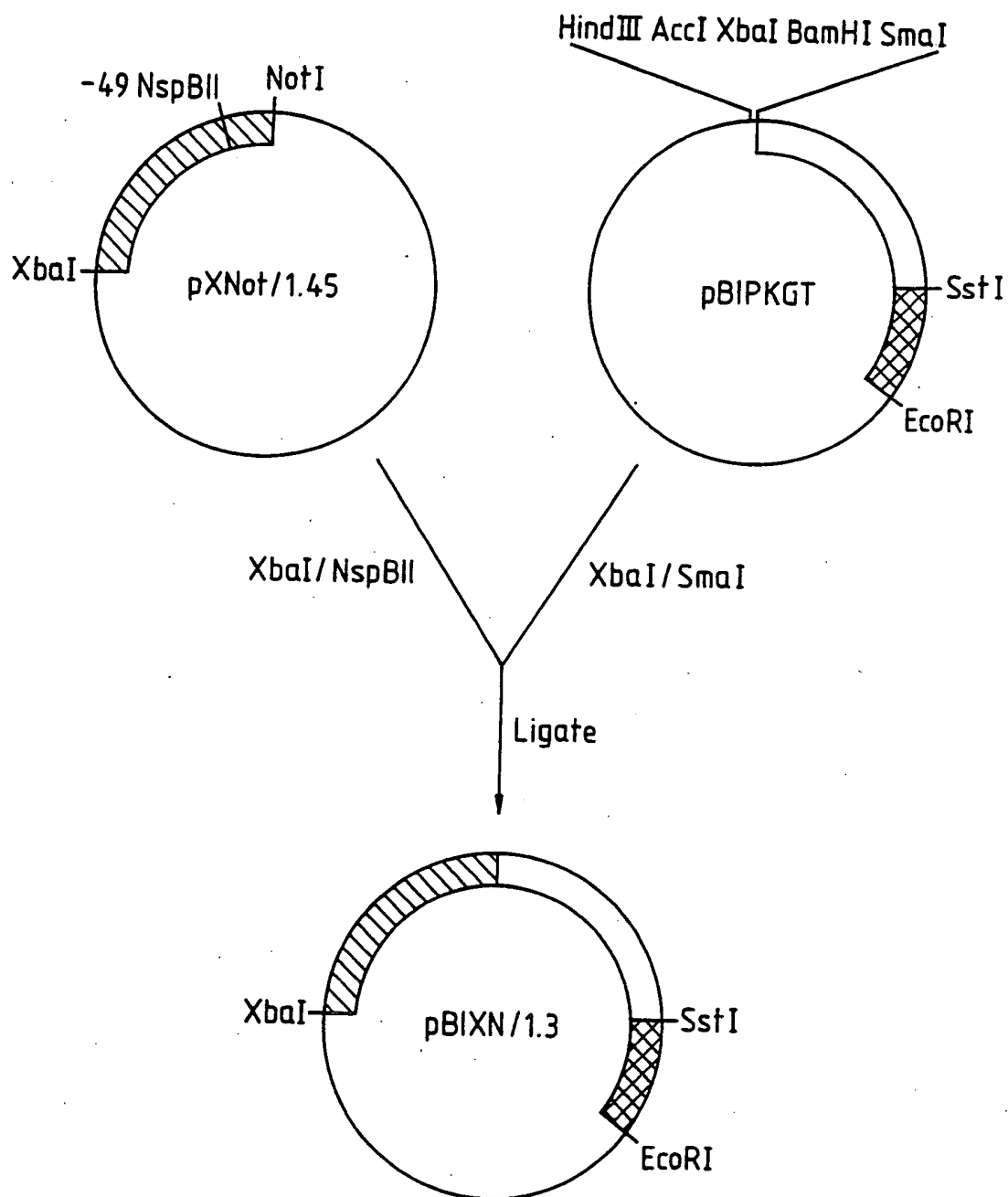
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Fig. 7.



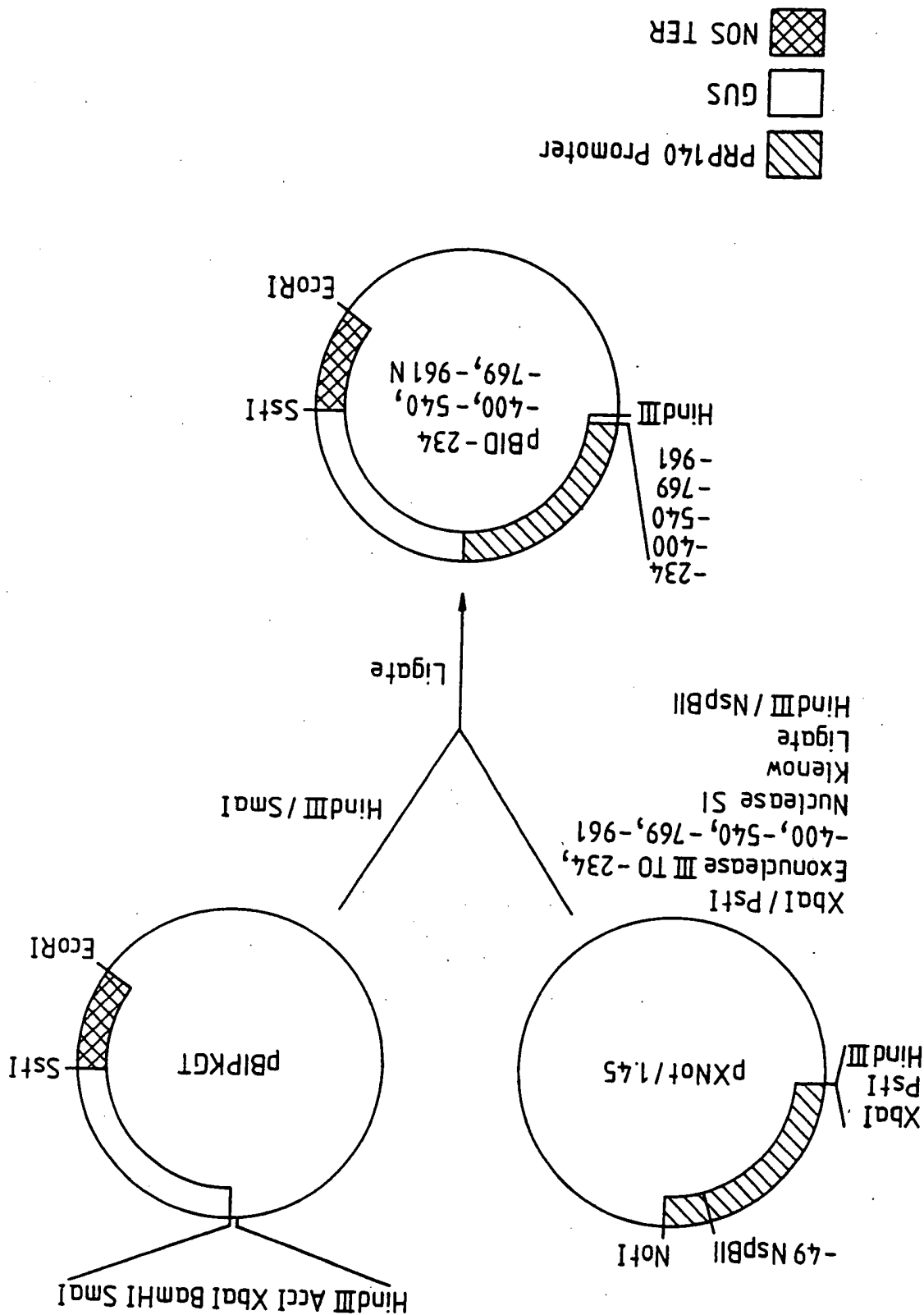
9/20

Fig. 8.



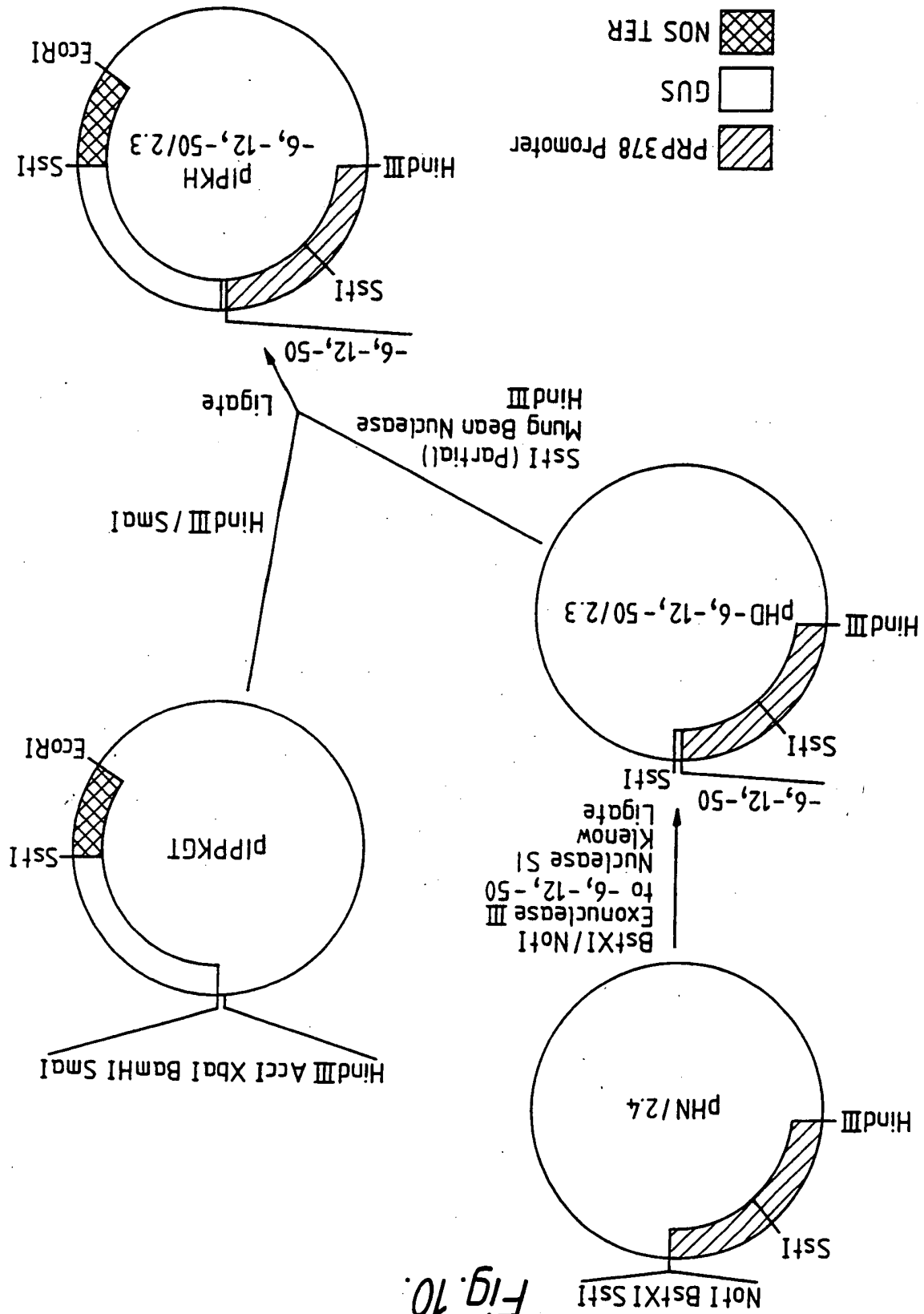
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Fig. 9.



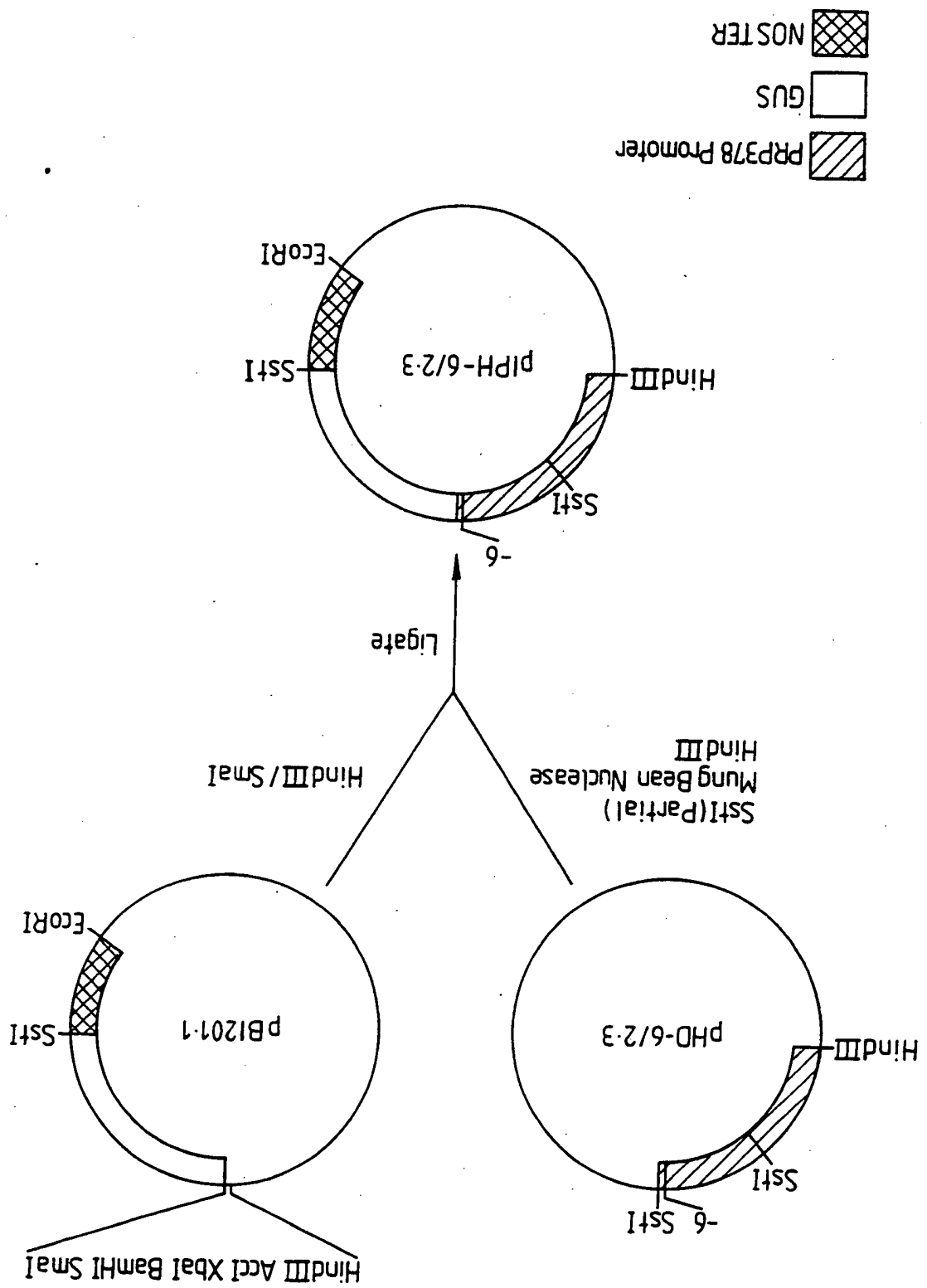
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Fig. 10.



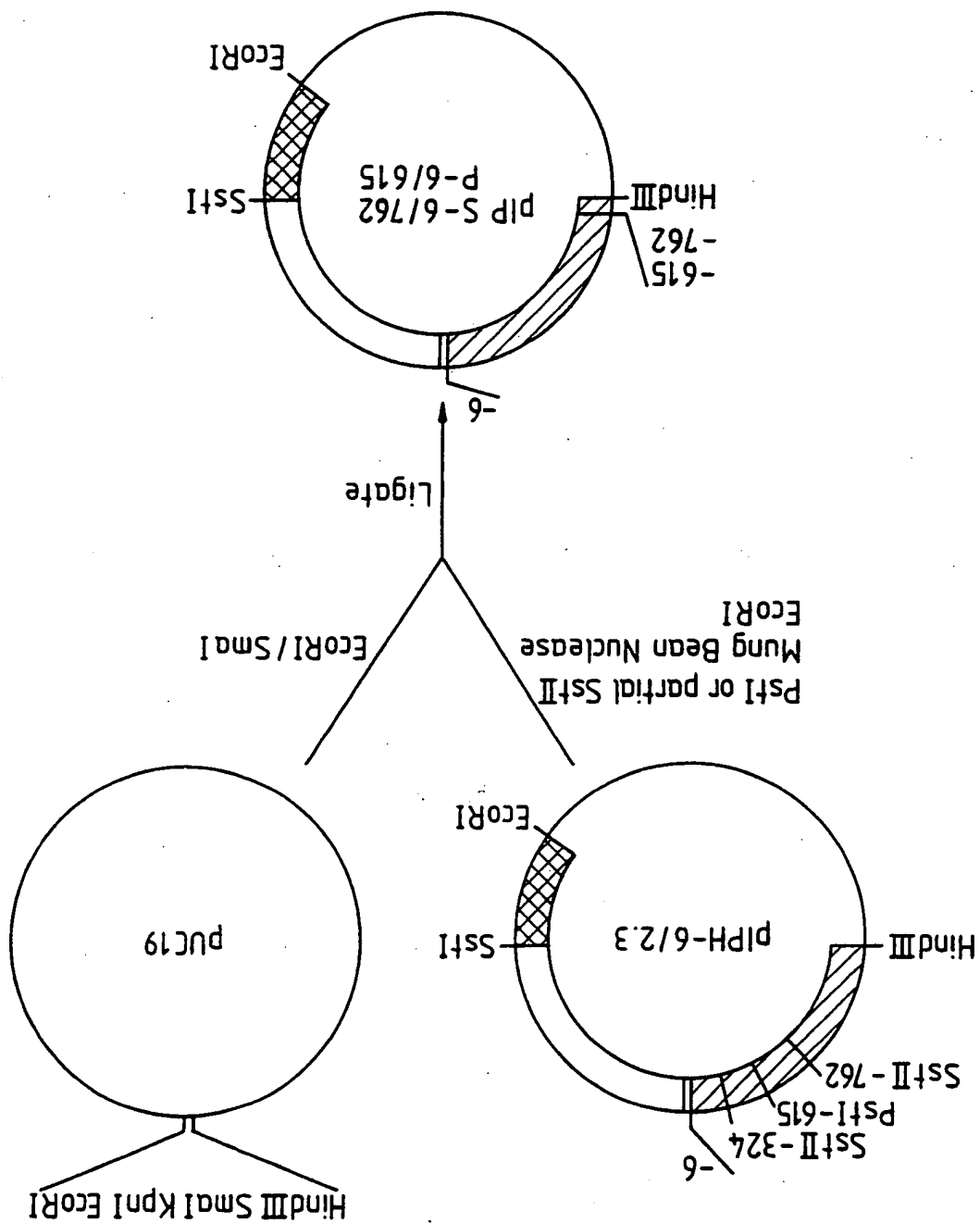
12/20

Fig. 11.



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Fig. 12.



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Fig.13.

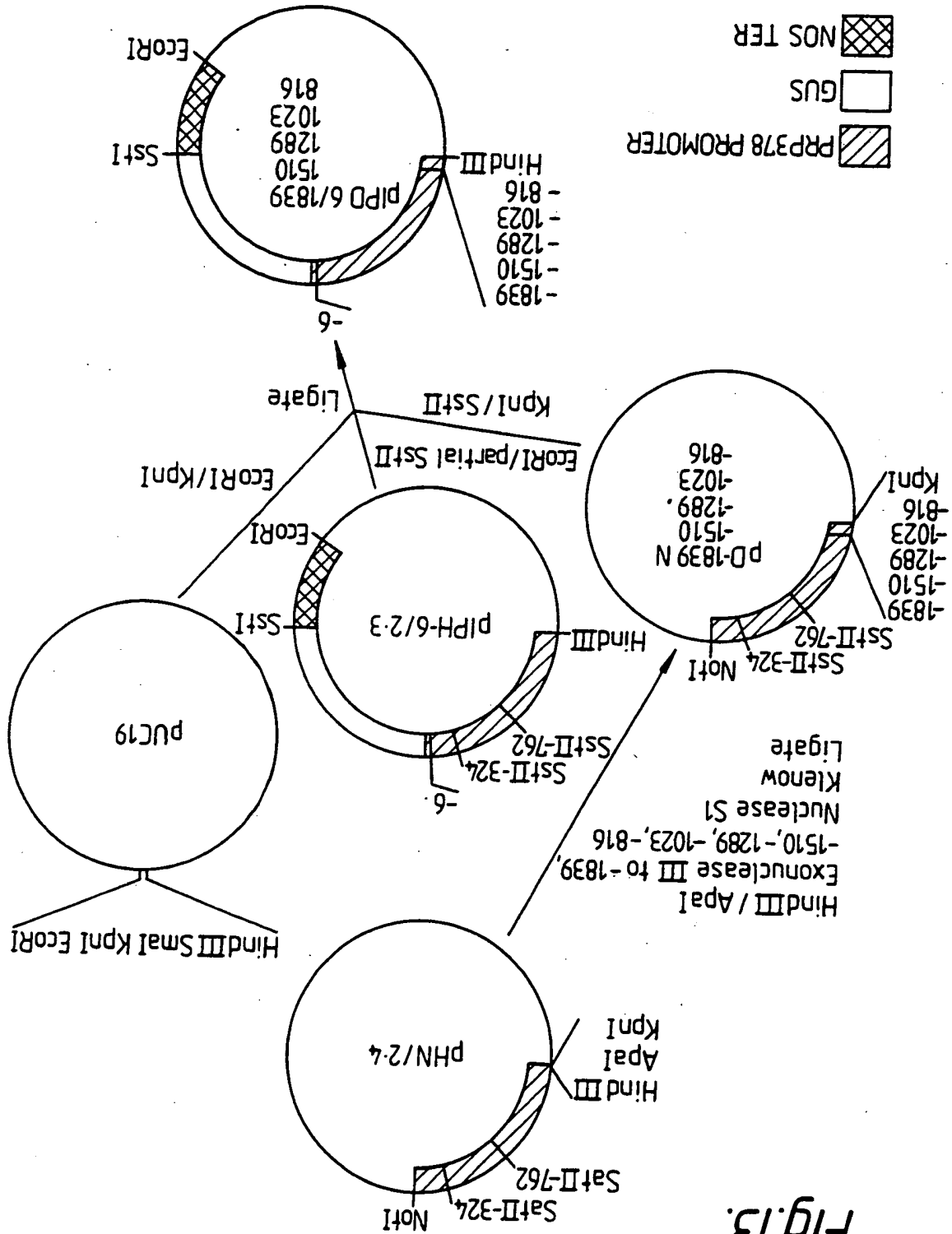
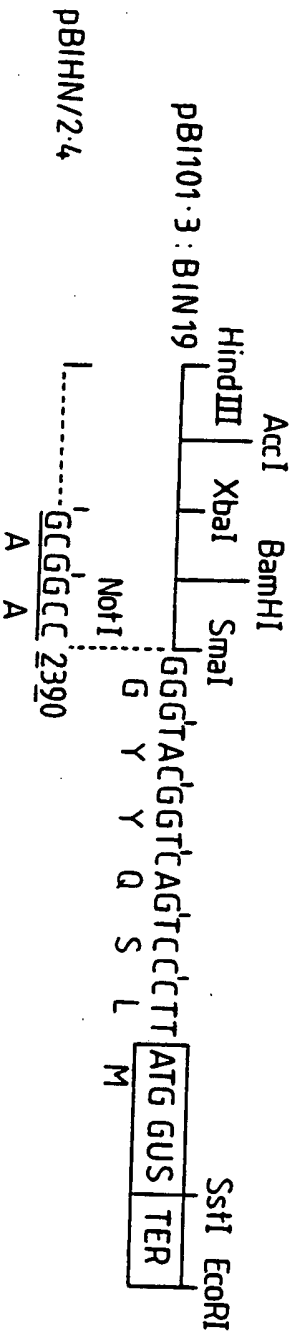
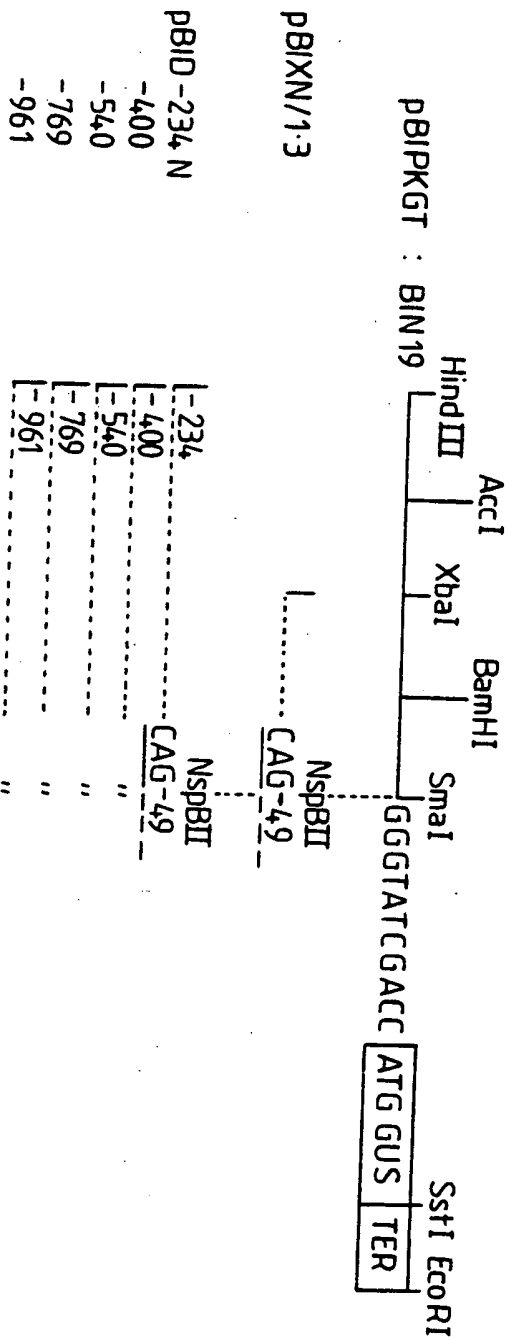


Fig. 14.

CONSTRUCT BASED ON:-



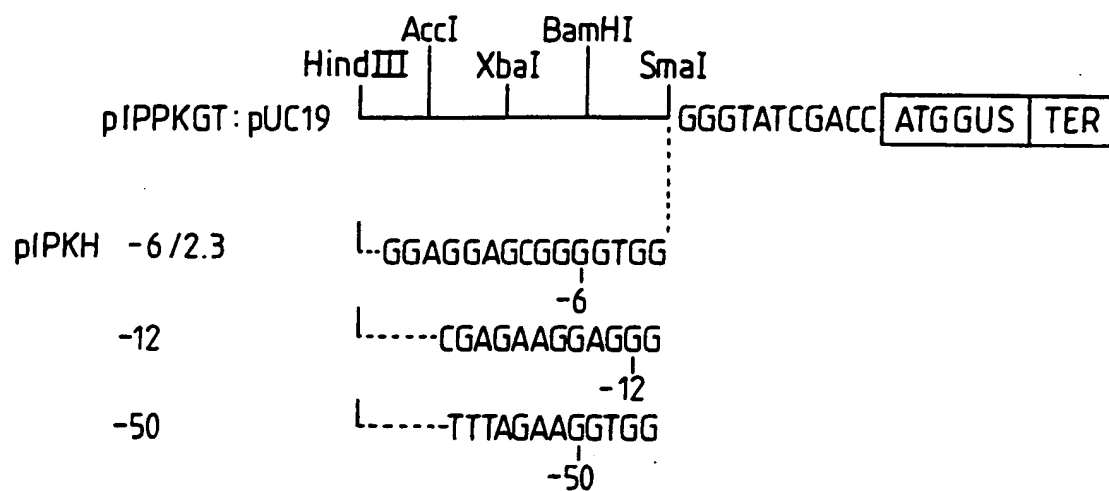
CONSTRUCTS BASED ON:-



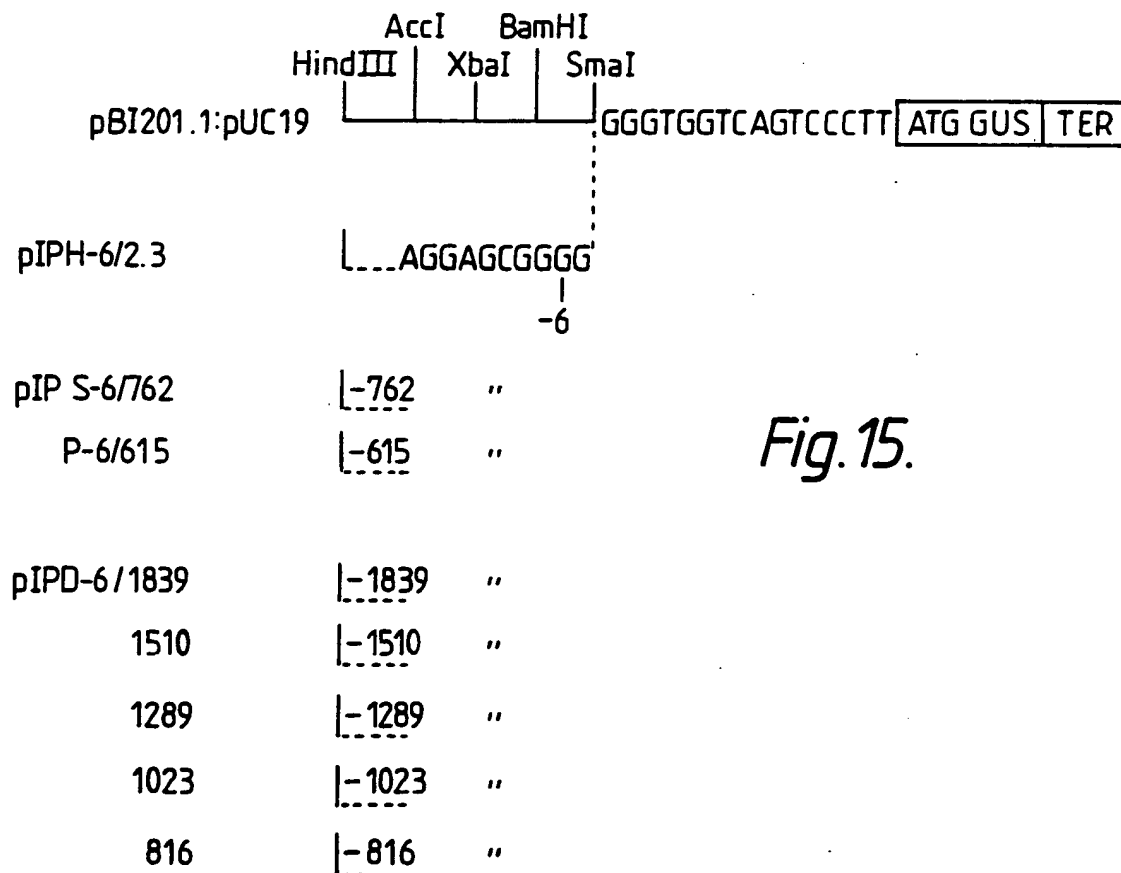
SUBSTITUTE SHEET

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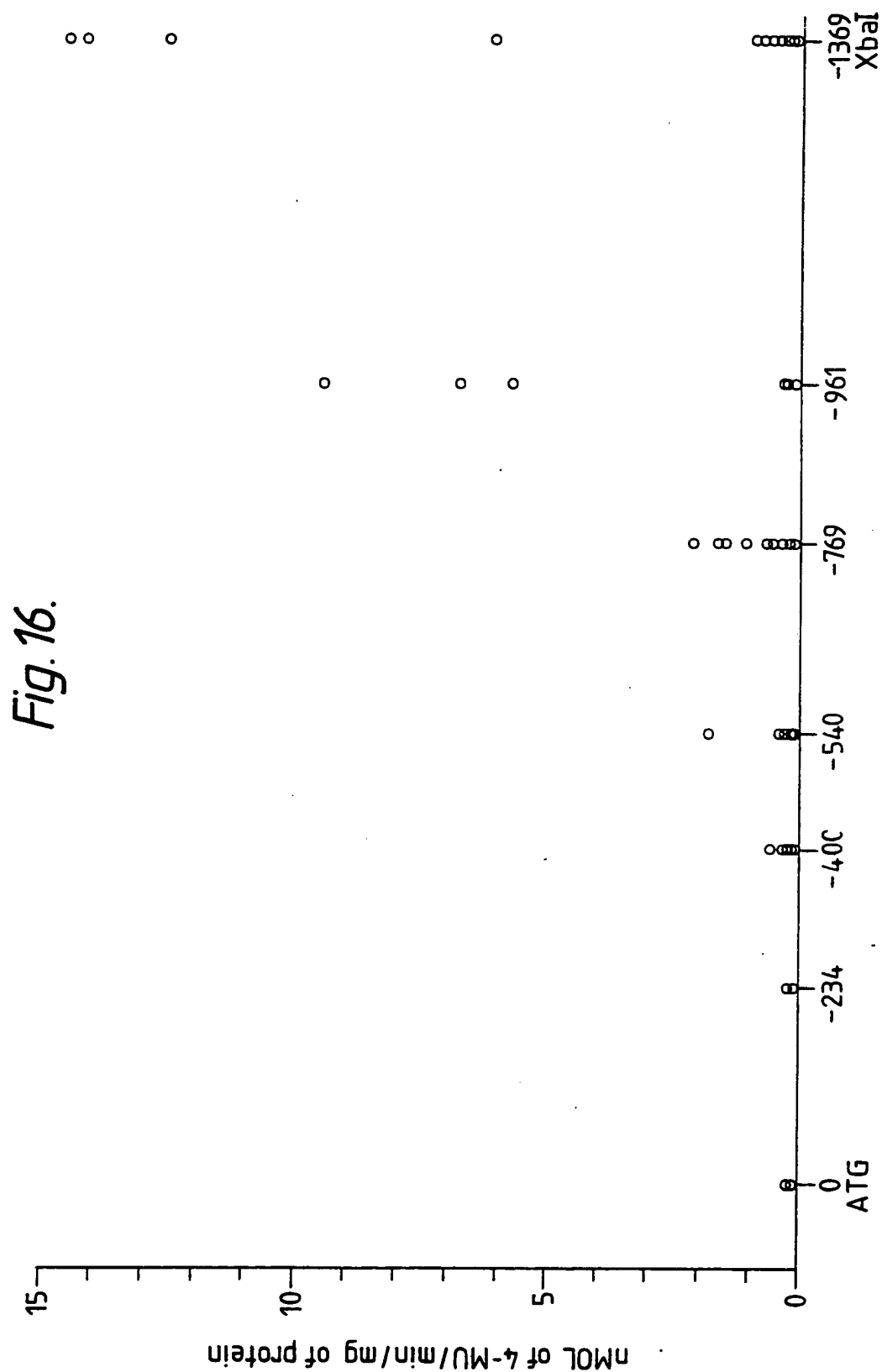
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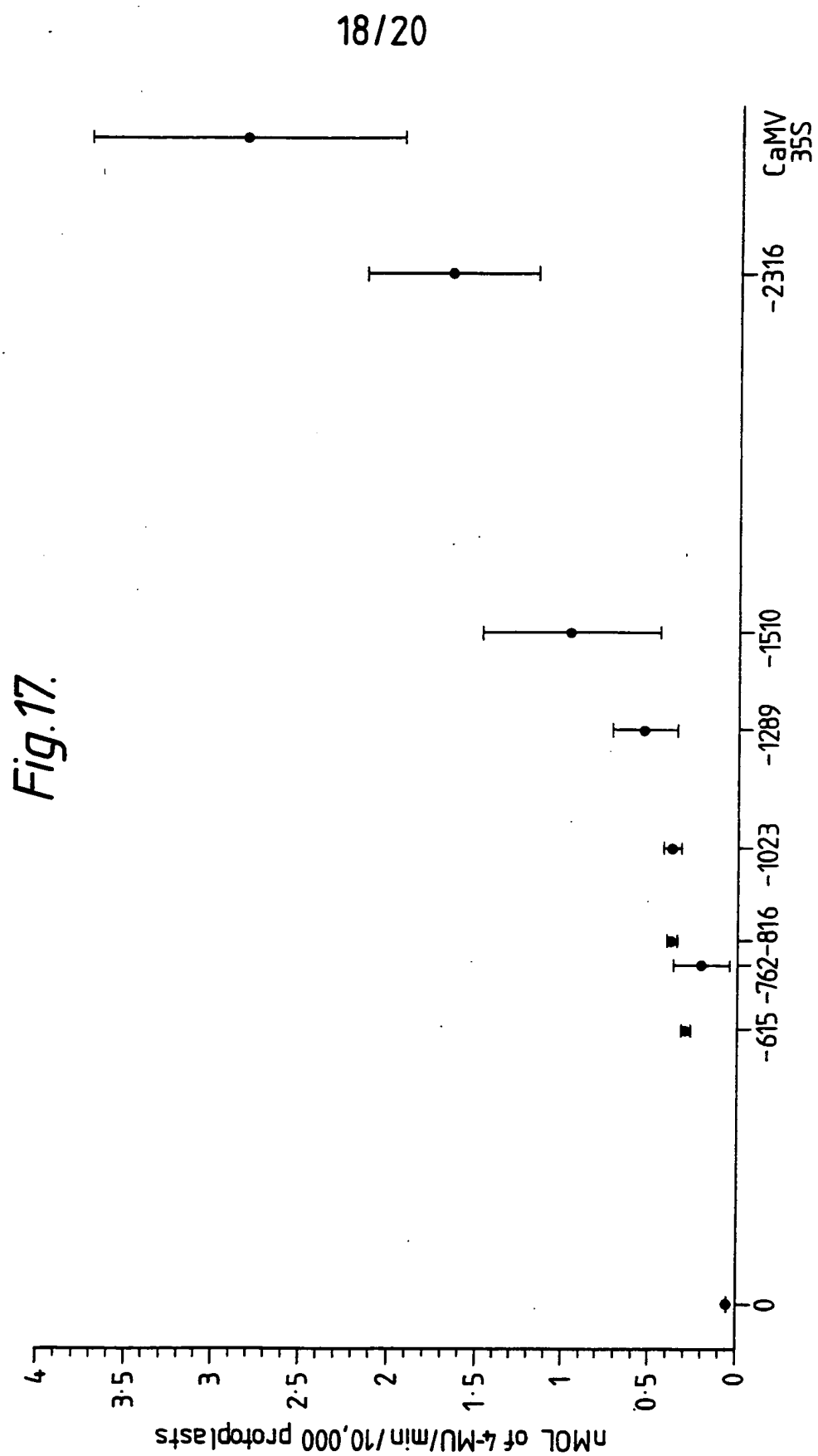


CONSTRUCTS BASED ON:

*Fig. 15.*

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Fig. 18 (cont.)

	961	CAAAGCCGGAAACCAAGCCAGAGCCCAAAACCGAGGCCCAAGCCAGAGCCGATGCCAAAAGCCGGAACCAAGCCGAGCCCC K P E P K P E P K P E P K P E P M P K P E P K P E P K P E P	1040
	1041	AAGCCCGAGCCAATGCCGAAGCCAGAACCGAAGCCAGAGCCCAAGCCTGAGCCGATGCCAAAAGCCAGAGCCTAAGCCCCAA K P E P M P K P E P K P E P K P E P K P E P M P K P E P K P E P K P K	1120
	1121	ACCATTCGCCTAAACCAGAGCCCTAAGCCTGAACCTATGCCTAAGCCAGAGCCCAAGCCTGAGCCCGAAACCGAAGCCGGAGC .P L P K P E P K P E P M P K P E P K P E P K P E P E P K P E P	1200
	1201	CGCCTCCGAAGGGCAAGCCACCGATGACTGACAATTGTATGTATACTCACATATGCAGCTGAAGGAGGAGATCGACCCCC P P K G K P P M T D N	1280
	1281	GTCCGGAGCCACGATGTGCTATTTCTAGAATAAGTGGCATAGTATCCGGTTAGCGAGATAGTGATGATGCATCTTTTTGT -----+-----+-----+-----+-----+-----+-----+-----+-----+	1360
	1361	ATTCCCTTGTAATCCACCATTCCTTTTAGTTTCTGTGTCTCCATGCACCCATGATGAGTACTACTATTCCTGTAATATATCAT -----+-----+-----+-----+-----+-----+-----+-----+-----+	1440
	1441	TTGCGTGTGGTATATGTTTCATCTGTGCACATGACTCAGTTGTTCTTTTCGTGTAGATACACATGCTTTTATTTCGTTTGCA -----+-----+-----+-----+-----+-----+-----+-----+-----+	1520
	1521	TGAAAGACACATTTTGTCCACCGAATTC-----+-----+-----+-----+-----+-----+-----+-----+----- -----+-----+-----+-----+-----+-----+-----+-----+-----	1548

INTERNATIONAL SEARCH REPORT

International Application No

PCT/GB 91/00397

I. CLASSIFICATION F SUBJECT MATTER (If several classification symbols apply, indicate all) *

According to International Patent Classification (IPC) or to both National Classification and IPC

IPC⁵: C 12 N 15/82, C 12 N 15/29, C 12 N 5/10, A 01 H 5/00

II. FIELDS SEARCHED

Minimum Documentation Searched *

Classification System

Classification Symbols

IPC⁵

C 12 N, A 01 H

Documentation Searched other than Minimum Documentation
to the extent that such Documents are included in the Fields Searched *

III. DOCUMENTS CONSIDERED TO BE RELEVANT *

Category *	Citation of Document, ¹¹ with indication, where appropriate, of the relevant passages ¹²	Relevant to Claim No. ¹³
------------	--	-------------------------------------

A

Plant Molecular Biology, volume 11, 1988
Kluwer Academic Publishers, (Dordrecht
NL),
V. Stiefel et al.: "Molecular cloning
of cDNAs encoding a putative cell wall
protein from Zea mays and immunologi-
cal identification of related polypep-
tides", pages 483-493
see the whole document

1-14

A

Journal of Cellular Biochemistry, Suppl.
13D, 1-7 April 1987, Alan R. Liss,
Inc., (New York, US),
M.P. Vallés et al.: "Structure and
expression of two families of develop-
mentally regulated genes from Zea
mays", page 286
see abstract M241

1-14

./.

* Special categories of cited documents: ¹⁴

"A" document defining the general state of the art which is not
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"E" earlier document but published on or after the international
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other means

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"T" later document published after the international filing date
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cited to understand the principle or theory underlying the
invention

"X" document of particular relevance; the claimed invention
cannot be considered novel or cannot be considered to
involve an inventive step

"Y" document of particular relevance; the claimed invention
cannot be considered to involve an inventive step when the
document is combined with one or more other such docu-
ments, such combination being obvious to a person skilled
in the art.

"A" document member of the same patent family

IV. CERTIFICATION*

Date of the Actual Completion of the International Search

14th June 1991

Date of Mailing of this International Search Report

19. 07. 91

International Searching Authority

EUROPEAN PATENT OFFICE

Signature of Authorized Officer

D. Frank

Mme Dagmar FRANK

III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)

Category *	Citation of Document, " with indication, where appropriate, of the relevant passages	Relevant to Claim No.
A	<p>Chemical Abstract, volume 112, 1990, (Columbus, Ohio, US), Keller, Beat et al.: "Specific expres- sion of a novel cell wall hydroxy- proline-rich glycoprotein gene in lateral root initiation", see page 209, abstract no. 113287y & Genes Dev. 1989, 3(10), 1639-46</p> <p>-----</p>	1-14